

complexes with elastase, i.e., productive ones able to transform into irreversibly inhibited enzyme and nonproductive ones unable to undergo further reaction. This explains unambiguously why the TFA-peptide-CMK possess higher affinities and lower rate constants than the corresponding acetyl-peptide-CMK (Table I).

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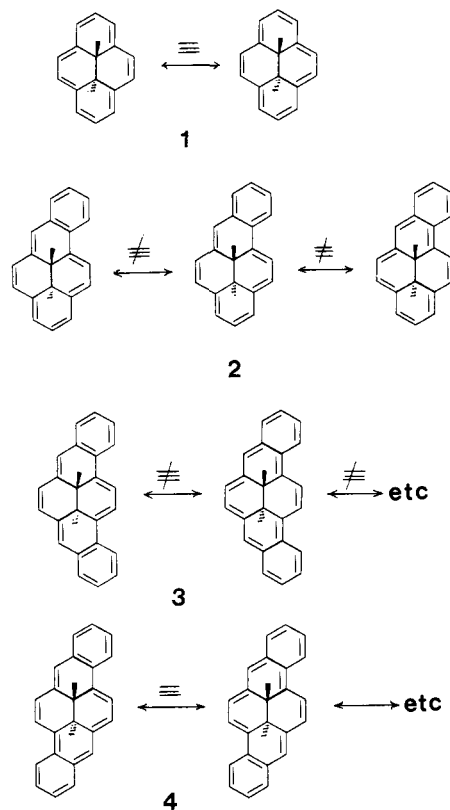
## A Clear Demonstration of Importance of Symmetrical Kekulé Structures to Diatropicity (Aromaticity). Synthesis of a Stable, Highly Diatropic, Dibenzannulene<sup>1</sup>

Sir:

Although the concept of aromaticity is often introduced by consideration of the two identical Kekulé structures of benzene, when the aromaticity of larger annulenes is discussed (usually by comparison of diatropicities), such structures are normally neglected. We present here a clear experimental demonstration

that, all other things being equal, the equivalence of Kekulé structures can have a profound effect on the diatropicity of an annulene. Nakagawa<sup>2</sup> has obtained results in the [18]annulene series which in our opinion also support the hypothesis, even though he suggests that his more recent<sup>3</sup> results throw doubt on his initial conclusions. This may be, however, because he is not comparing geometrically equivalent systems, or because of the inclusion of cumulated bonds. We believe that Boekelheide's<sup>4</sup> *trans*-15,16-dimethyldihydropyrene<sup>5</sup> (**1**) with its planar,<sup>6</sup> rigid [4n + 2]  $\pi$ -electron periphery, and easily discernable, highly shielded ( $\delta$  -4.25) internal methyl protons is an excellent system to study to detect any such effect.

We thus now report the synthesis and properties of the mono- and bisbenzannulated derivatives of **1**, namely **2**, **3**, and **4**.



Reaction of 1,3-bis(bromomethyl)-2-methylnaphthalene,<sup>7a,8</sup> respectively, with 2,6-bis(mercaptomethyl)toluene<sup>7b</sup> and 1,3-bis(mercaptomethyl)-2-methylnaphthalene<sup>8</sup> (mp 88–89 °C prepared by the thiourea method<sup>7b</sup>) under high dilution conditions yielded the thiacyclophanes **5** (mp 188–190 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 60 MHz)  $\delta$  8.3–7.0 (m, 8 H, ArH), 4.16, 4.13, 3.75, 3.70, 3.40, 3.35 (all s, 1 H, 1 H, 2 H, 2 H, 1 H, 1 H, -CH<sub>2</sub>-), 1.42 (s, 3 H, -Np-CH<sub>3</sub>), and 0.92 (s, 3 H, -PhCH<sub>3</sub>))

Table I

Compd	Color of crystals	Mp, °C	NMR spectra				
			External H, $\delta$	External C, ppm	Internal bridge C, ppm	Internal methyl H, $\delta$	Internal methyl C, ppm
<b>1</b> <sup>a</sup>	Green	115–116	8.7–8.1	137–123	30.0	-4.25 <sup>b</sup>	14.0
<b>2</b>	Orange-purple	218–220	8.7–7.1	139–117	35.5, 36.0	-1.60 <sup>b</sup>	17.0, 17.7
<b>3</b>	Green	195–196.5	8.2–6.9	139–117	39.5	0.02 <sup>b</sup>	19.2
<b>4</b>	Blue	195–196.5	9.8–7.8	137–124	32.8	-3.58 <sup>c</sup>	15.9 <sup>d</sup>

<sup>a</sup> R. DuVernet and V. Boekelheide, *Proc. Natl. Acad. Sci. U.S.A.*, **71**, 2961 (1974). <sup>b</sup> CDCl<sub>3</sub>. <sup>c</sup> THF-d<sub>8</sub>. <sup>d</sup> Tentative; this peak is very weak, possibly owing to relaxation time difficulties. The <sup>1</sup>H NMR spectra were recorded on a Perkin-Elmer R32 90-MHz spectrometer and <sup>13</sup>C NMR spectra in CDCl<sub>3</sub> on a Nicolet TT-14 60-MHz Fourier transform spectrometer, the chemical shifts being given in parts per million downfield from Me<sub>4</sub>Si.

